

Screening and Evaluation of Phytochemical Constituents, Antimicrobial, Antioxidant and Anti-Arthritic Activities of Methanolic Extract of *Nyctanthes arbortristis* L. Leaves

Tata V.S. Padmavathi¹ and Audipudi Amrutha V.^{2*}

1. Department of Botany, Government Degree College, Eluru, Andhra Pradesh, INDIA

2. Department of Botany and Microbiology, Acharya Nagarjuna University, Guntur-522510, Andhra Pradesh, INDIA

*audipudiamrita@gmail.com

Abstract

The objective of the current research was to determine the antioxidant, antimicrobial and anti-arthritic activities of the methanolic extract of *Nyctanthes arbortristis* leaves (MLEN). In the present study, the phytochemical constituents, total phenolic content and reducing power of MLEN were analysed and the antimicrobial activity of MLEN against three clinical pathogens: *Staphylococcus aureus* (MTCC 9542), *Fusobacterium nucleatum* (MTCC 25586) and *Propionibacterium acnes* (MTCC 1951) was studied using the agar-well diffusion method. We evaluated the antioxidant characteristics and anti-arthritis activities using the DPPH assay and in vitro inhibition of egg albumin denaturation. MLEN showed the presence of phenols, tannins, cardiac glycosides, alkaloids and flavonoids, with a total phenolic content equivalent to 450 µg/ml of gallic acid and a maximum reducing power (0.88) at 80 µg/ml.

The leaf extract showed 28.28 mm/0.1 ml inhibition with *S. aureus*, 25.75 mm/0.1 ml with *F. nucleatum* and 13.18 mm/0.1 ml with *P. acnes*. MLEN showed significant antioxidant activity (IC₅₀ 59.7 µg/ml). In vitro anti-arthritic activity, achieved through the inhibition of protein denaturation, yielded promising results with an IC₅₀ of 109.3458 µg/ml. Outcomes of the investigation revealed that phenols, tannins, cardiac glycosides, alkaloids and flavonoids may be the primary metabolites in MLEN responsible for significant antimicrobial, antioxidant and anti-arthritic activities, paving the way for further investigation of biologically active principles of pharmacological significance.

Keywords: *Nyctanthes arbortristis*, phytochemical screening, antimicrobial activity and antioxidant potential.

Introduction

Nowadays, pharmaceutical research on herbal medicine is gaining more attention due to its therapeutic use as an alternative to conventional medicine, as plants are a rich source of relatively harmless, potentially beneficial chemical compounds that can be used as drugs to treat various human

ailments^{1,4,14}. *Nyctanthes arbortristis* is one of the most popular plants in Ayurveda and herbal medicine with extensively wealthy phytochemical constituents, some of which are still to be explored.

N. arbortristis (night-flowering jasmine, coral jasmine and Parijat), which belongs to Nyctaginaceae, is a large shrub having a broad spectrum of pharmacological properties and is extensively cultivated in tropical and subtropical areas from India to Nepal, from the outer Himalayas and Jammu and Kashmir to east of Bengal, Tripura and Assam, up to the central part of the extended Godavari in the south^{5,8} and is commonly found in red as well as black soils with a 5.6 to 7.5 pH³⁰. Ayurveda, Siddha and Unani medicine systems used *N. arbortristis* as a laxative, digestive aid, diuretic, expectorant, antivenom and mild bitter tonic^{2,10,34,41}.

The phytochemical evaluation of *N. arbortristis* is gaining significance due to the increasing interest in natural products as an alternative source of medicine. It makes a significant contribution in expanding the knowledge of the chemical constituents of *N. arbortristis* and provides a scientific basis for its traditional uses. The exploration of plant-based bioactive secondary metabolites, their isolation, purification, characterisation and identification of specific bioactive compounds, will play a pivotal role in developing novel drugs or therapeutic agents. Several phytochemical constituents, including carbohydrates, steroids, flavonoids, iridoid glycosides, terpenes and alkaloids, available in different parts of *N. arbortristis*, exhibited varying pharmacological activities^{12,45}.

Quinolinolids, 7-(alpha-anilino-p-nitro benzyl)-8-quinolinol, protoberberines and aporphines reported in the leaves of *N. arbortristis* exhibited anti-inflammatory properties¹⁹. Methanolic stem bark extract of *N. arbortristis* exhibits anti-inflammatory properties, as was reported in Wistar albino rats (assessed by carrageenan-induced rat paw oedema)¹⁸. Similarly, seed extracts demonstrated potential antioxidants^{26,31}, antibacterial properties^{3,6,43}. Polyphenol extract of *N. arbortristis* leaves has been reported to contain gallic acid, chlorogenic acid, protocatechuic acid and caffeic acid, exhibiting effective free radical scavenging activity²⁴. This current research aimed to demonstrate the antimicrobial and antioxidant activities of bioactive phytochemical constituents present in the methanolic leaf extract of *N. arbortristis* to identify lead molecules of pharmacological significance.

Material and Methods

Collection of plant material and authentication: The collection of *N. arbortristis* leaves was carried out in distinct areas of the Eluru district, Andhra Pradesh, India. Prof. S. M. Khasim, Department of Botany and Microbiology, Acharya Nagarjuna University, Guntur 522510, Andhra Pradesh, India, authenticated the plant's taxonomic identity.

Extraction of the plant material: Fresh leaves from the *N. arbortristis* plant were washed, dried at room temperature and then crushed into a powder using a mechanical grinder. The leaf extract was prepared by mixing 258 g of leaf powder with 3100 mL of methanol. Methanolic extract (MLEN) was prepared at 35-40°C in the Soxhlet apparatus and concentrated to 1 mg/ml (W/V) under reduced pressure using a rotary evaporator and stored at 4°C until use⁴.

Chemicals and Clinical Pathogens: Clinical pathogens (*Staphylococcus aureus*-MTCC 9542, *Fusobacterium nucleatum*-MTCC 25586 and *Propionibacterium acnes*-MTCC 1951), Mayer's reagent, cell culture medium NAM, DPPH, 10 ml serological pipettes (TORSON), T25 flask (#12556009, Biolite-Thermo) and 50 ml centrifuge tubes (#54 043 TORSON) were used. Methanol was purchased from Sigma-Aldrich Co., USA.

Equipment: Soxhlet apparatus (Borosil), Rotary Evaporator (Heidolph, Germany), Centrifuge (Remi: R-80C), Hot Air Oven (Healforce, China), Incubators (Healforce, China), Pipettes: 2-10 µL and 10-100 µL, UV-Spectrophotometer were used.

Phytochemical constituents: A phytochemical screening of the extract was carried out using various tests to detect the compounds. Carbohydrates by Molisch's test, alkaloids by Mayer's test, glycosides by the foam test for saponin glycosides and Bontrager's test for both cardiac glycosides and anthraquinon glycosides. Flavonoids were tested with the lead acetate test and tannins were identified using the ferric chloride test. Terpenoids were assessed using the salkowski test, while fats and oils were screened with the saponification test. Coumarins were detected using the fluorescence test and proteins were identified using biuret's test⁴.

Quantitative Analysis of Total Phenolic Content [TPC]:

Total phenolic content of extracts was determined as described. One ml of extract (1 µg/ml) was added to 1 ml of Folin-Ciocalteu's reagent and saturated with Na₂CO₃ for 3 minutes. The mixture was diluted to a final volume of 10 ml and incubated for an hour in the dark. Absorbance was measured at 725 nm. Gallic acid was taken as the standard.

Estimation of Reducing Power: Different concentrations of the extract were added to 1 mL of distilled water and 1 mL of 0.2 M phosphate buffer, along with 1% (K₃[Fe (CN)₆]). The mixture was incubated for 20 min at 50°C. To this, 1 mL of 100 mg/mL trichloroacetic acid was added and

the mixture was centrifuged for 10 minutes at 8000 × g. Distilled water and FeCl₃ were added to the upper layer of solution. At a wavelength of 700 nm, absorbance was measured. As a standard reference, ascorbic acid was used.

Antimicrobial activity: The antimicrobial activity of the plant extract has been studied using the agar well diffusion method, with methanol as the control. Three bacterial strains, namely *Staphylococcus aureus* (MTCC 9542), *Fusobacterium nucleatum* (MTCC 25586) and *Propionibacterium acnes* (MTCC 1951), were obtained from MTCC, Chhattisgarh and cultured in nutrient agar medium. The streaking technique was used to inoculate the bacterial culture with a concentration of 10⁸ CFU. 100 µL and 200 µL of MLEN and 100 µL of methanol, were separately poured into wells with an 8 mm diameter and incubated for 24 to 48 hours at 37°C. The diameter of inhibitory zones surrounding each well was measured in mm following incubation. Methanol served as a negative control. Streptomycin served as a positive control. Zones of inhibition were measured by measuring their diameters.

DPPH free radical scavenging assay: DPPH was used to assess MLEN's scavenging capacity for free radicals. 3 mL of MLEN was prepared in different concentrations (20, 40, 60, 80, 100 µg/mL). 1 mL of 0.004% methanolic DPPH was added to each concentration and placed in the dark for 30 minutes. Ascorbic acid was used as a standard and the absorbance was measured at 517 nm. The % inhibition was calculated using the formula:

$$\% \text{ inhibition} = \frac{[\text{absorbance of control} - \text{Absorbance of sample}]}{\text{Absorbance of control}} \times 100.$$

In vitro anti-arthritis activity: The *in vitro* anti-arthritis property was determined by the inhibition of egg albumen denaturation. 5-ml reaction mixture is composed of 0.2 ml egg albumin, 2.8 ml phosphate-buffered saline and 2 ml MLEN and is incubated at 37°C for 15 minutes, followed by incubation at 70°C for 5 minutes. The effective dose was determined using extracts at varying concentrations (20, 40, 60, 80, 100 µg/ml), with 5 ml of double-distilled water as the control. Absorbance was measured at 660 nm at room temperature. Various doses of diclofenac sodium (20, 40, 60, 80 and 100 µg/ml) were taken as the reference medication. The formula for calculating the % inhibition of protein denaturation is:

$$\% \text{ of inhibition} = \frac{[V_c - V_t/V_c]}{V_c} \times 100$$

where V_c is absorbance of control and V_t is Absorbance of test sample

Results and Discussion

Phytochemical screening: Research on medicinal plants for their chemical constituents has become increasingly important due to their therapeutic value and potential. As part of the current investigation, a crude methanolic extract

of *N. arbortristis* leaves has been analysed for its chemical constituents. As reported earlier^{7,16,22,33,37,38,45,48}, MLEN also contained alkaloids, flavonoids, glycosides, terpenoids, tannins and saponins (Table 1). In the essential oil of *N. arbortristis*, compounds such as geranylgeraniol, cis-9-tricosene, benzyl salicylate, n-pentacosane, phytone, nonadecane and methyl stearate were found to be present while the anthraquinone compound was absent⁴². The metal analysis studies on *N. arbortristis* revealed the presence of heavy metals at permissible levels, indicating that the plant was safe for use in herbal drug formulations⁴⁶.

Quantitative Analysis of Total Phenolic Content [TPC]:

Phenols present in the plant extract reacted with phosphomolybdenum and produced a blue colour. The plant extract exhibits an absorbance of 1.47 at 725 nm, equivalent to 450 µg/ml of gallic acid, as determined by the Folin and Ciocalteu method.

Estimation of reducing power: When estimating the reducing power of plant extracts, the absorbance values at 700 nm increased as the concentration of the plant extract increased. The maximum absorbance was recorded at 0.88 at a concentration of 80 µg/ml. It coincided with the concentration of the highest antioxidant activity, which is 80 µg/ml, indicating the optimum concentration of plant extract for antioxidant activity (Figure 1).

Antimicrobial activity: Three bacterial strains were employed to assess the plant extract's antibacterial properties using the agar well diffusion method, namely *S. aureus* (MTCC 9542), *F. nucleatum* (MTCC 25586) and *P. acnes* (MTCC 1951). The zones of inhibition at 100 µL concentrations for these bacteria are 4.47 ± 0.40 mm, 4.09 ± 0.40 mm and 2.11 ± 0.40 mm respectively. At a 200 µL concentration, the zones of inhibition are 5.66 ± 0.40 mm, 4.41 ± 0.40 mm and 3.65 ± 0.40 mm respectively. The results indicated that plant extract is most effective on *S. aureus* (MTCC 9542), moderately effective on *F. nucleatum* (MTCC 25586) and least effective on *P. acnes* (MTCC 1951) (Fig. 2 and Table 2).

The aqueous extracts of dried leaves demonstrated anti-malarial activity in an experimental study on patients suffering from malaria in Worli, Mumbai¹³. Ethanolic, methanolic and chloroform leaf extracts showed a marked antibacterial activity against different pathogenic bacteria including *S. aureus*, *P. acnes*, *Pseudomonas aeruginosa* and *Salmonella typhi*^{10,32,40}. With an inhibition zone diameter of 23.8 as well as 26.3 mm at 1000 µg/ml in opposition to *Klebsiella pneumoniae* and *Pseudomonas aeruginosa* respectively, essential oil eugenol and its derivatives from dried leaf extracts of *N. arbortristis* had the most potent antibacterial activity²⁰. Ethyl acetate and dichloromethane extracts of *N. arbortristis* exhibited good antioxidant activity against *B. subtilis*, *B. cereus* and *Pseudomonas*¹⁵. *N. arbortristis* plants have excellent antimicrobial activity²⁹. According to the results above, the antimicrobial activity of

each bacterial species increases as the concentration of the plant extract rises. The maximum activity was observed with *S. aureus* while it showed the least effect on *P. acnes* at both concentrations. Inhibition of *F. nucleatum* is moderate compared to the other two species.

Antioxidant activity: A DPPH assay was used to quantify MLEN's capacity to scavenge free radicals. The inhibition percentage gradually increased with an increase in the concentration of the plant extract from 20 to 80 µg/ml and then declined at 100 µg/ml. Maximum % of inhibition was observed at 80 µg/ml with an IC₅₀ value of 59.7 µg/ml, indicating potent antioxidant activity of plant extract (Fig. 3). This result was supported by a study by Michael et al²⁵ who displayed an IC₅₀ value of 57.93 µg/ml with MLEN. High antioxidant activity has been proved with methanol³⁹ and ethyl acetate extracts of root⁹, stem and flower of *N. arbortristis*, with excellent scavenging action of aqueous extract of leaf¹⁷.

In rats induced with diabetes by streptozotocin, Rathod et al³⁶ demonstrated the ability to scavenge free radicals. Hydroethanolic leaf extract of *N. arborists* showed good free radical scavenging activity on the liver of a Freund's adjuvant-induced arthritis model²³. Devasree et al¹¹ successfully noted the impact of *N. arbortristis* leaf aqueous extract on the immunological response of *Oreochromis mosambicus*. The ethanolic leaf extract of *N. arbortristis* showed high antioxidant property in high-fat streptozotocin diet-induced diabetic rats²⁷. Significant antioxidant activity of ethyl acetate extract from the leaf was noticed because of its scavenging ability of ROS, DNA strand breaks and lipid peroxidation³⁵ and antioxidant activity in different solvent extracts of flowers²¹. The antioxidant activity was higher in the calyx than in the petals of the aqueous flower⁴⁷. Extracts of *N. arbortristis* showed potential antioxidant activity⁴⁴. Comparative analysis of the antioxidant activity of a plant's roots, flowers, leaves and seeds with successive extracts, methanolic and aqueous extracts showed more antioxidant capacity than petroleum ether and chloroform extracts. The free radical scavenging activity is attributed to various bioactive compounds such as polyphenols, present in the plant parts¹⁷.

Anti-arthritic activity: Comparing MLEN to the standard medication diclofenac, the current investigation showed a significant decrease in protein denaturation. Protein denaturation inhibition % increased with concentration, with an IC₅₀ of 109.3458 µg/ml. At 60 µg/ml, 80 µg/ml and 100 µg/ml, the plant extract's inhibition percentage is higher than that of diclofenac (Fig. 4). One of the main concerns of inflammation in arthritis is protein denaturation. In some arthritic diseases, protein denaturation, along with the action of proteases, may lead to the production of autoantigens which are characteristic of certain arthritic conditions⁴. Leukocyte proteinase causes tissue damage during inflammatory reactions and by inhibiting proteinase activity, protection is provided against inflammation³⁶.

Conclusion

The availability of phenols, carbohydrates, flavonoids, alkaloids, glycosides, terpenoids and saponins was revealed through the screening of the phytochemical elements of *Nyctanthes arbor-tristis*. *Staphylococcus aureus* is the strain

most responsive to the concentration-dependent inhibition of bacterial growth via methanolic leaf extract from *N. arbor-tristis*. It supports the possible application of *N. arbor-tristis* as a natural antimicrobial agent. Future investigations will be needed to optimise concentration and to explore its mechanism of action.

Table 1
Phytochemical constituents of MLEN

Phytochemicals	MLEN
Alkaloids	+
Flavonoids	+
Glycosides	+
Terpenoids	+
Tannins	+
Saponins	+
Anthraquinones	-

Table 2
Antimicrobial Activity of MLEN [Zone of Inhibition in mm]

Bacterial Strain	Zone of Inhibition [100µL]	Zone of Inhibition [200µL]	Circumference of inhibition mm/0.1ml
<i>S. aureus</i>	4.47± 0.40	5.66±0.40	28.28
<i>F. nucleatum</i>	4.09± 0.40	4.41±0.40	25.75
<i>P. acnes</i>	2.11 ±0.40	3.65 ± 0.40	13.18

Values are represented in Mean ±SD (n=3)

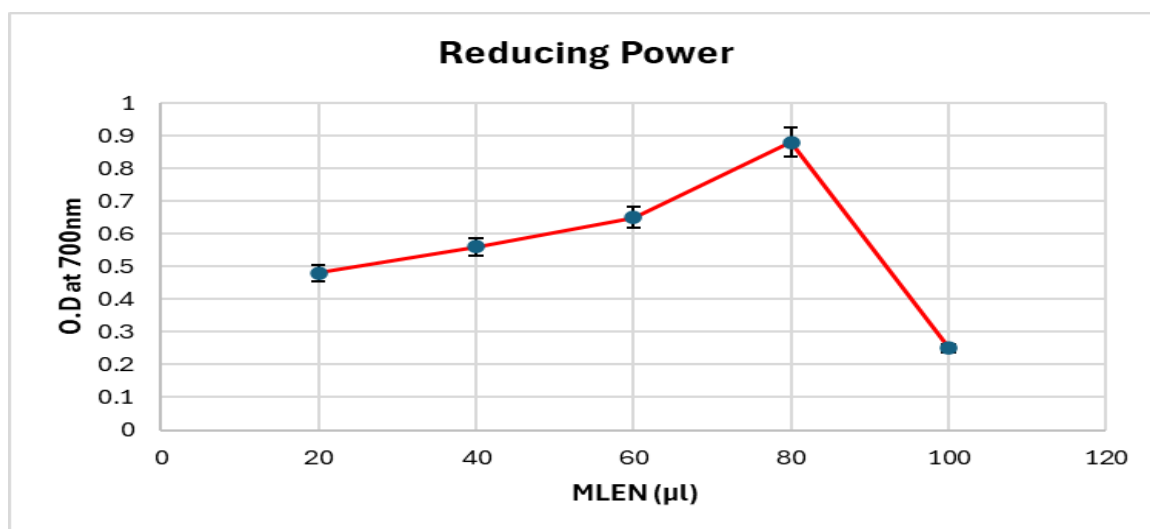


Fig. 1: Estimation of reducing power of MLEN

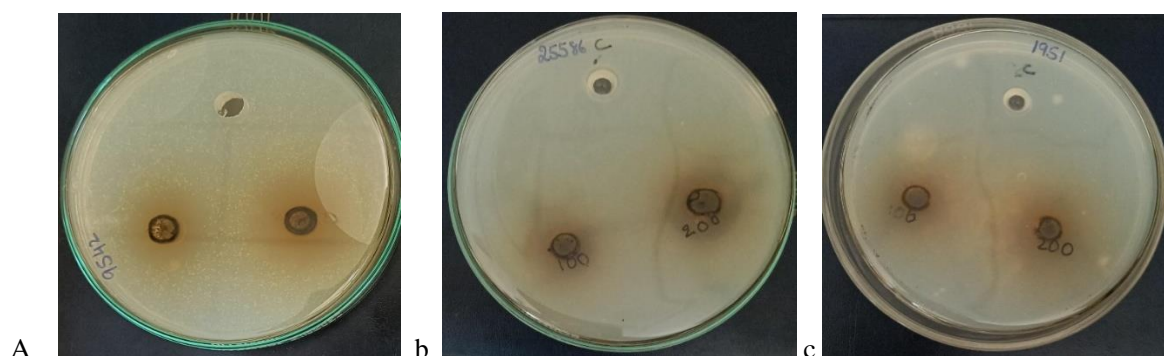


Fig. 2: Antimicrobial Activity of MLEN

[a] *Staphylococcus aureus* [b] *Fusobacterium nucleatum* and [c] *Propionibacterium acnes*

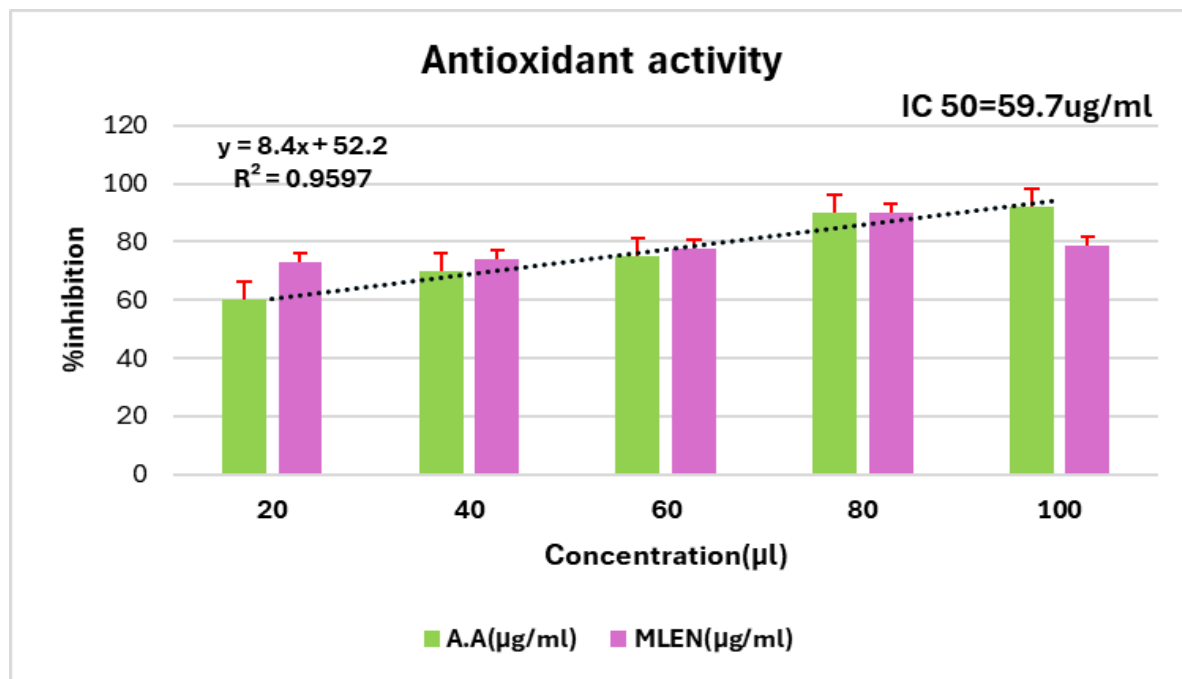


Fig. 3: Antioxidant activities of MLEN in comparison with Ascorbic acid

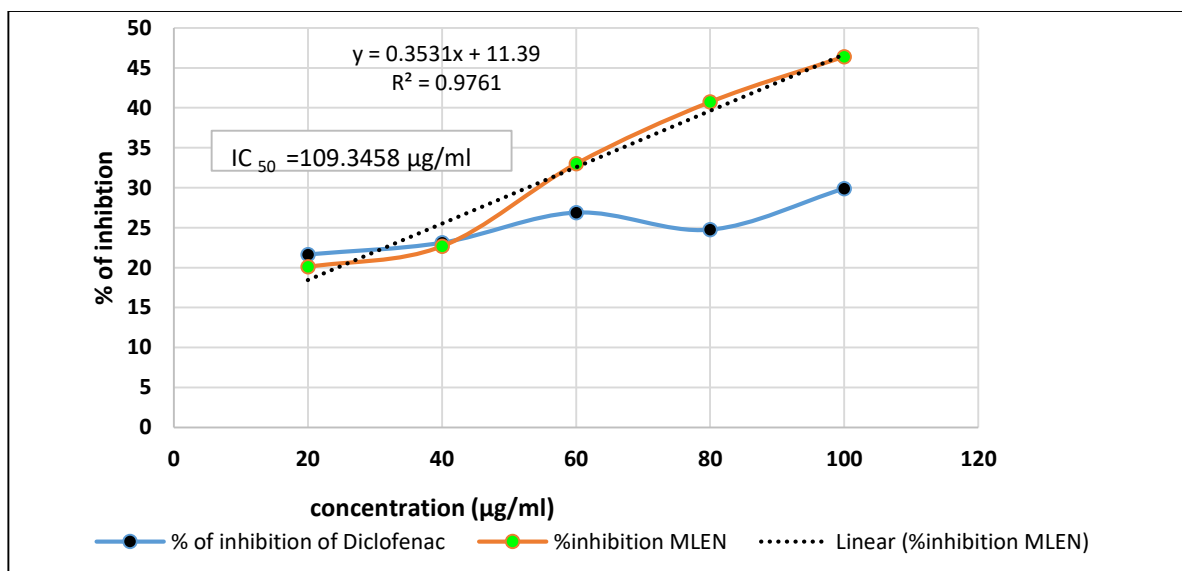


Fig. 4: Antiarthritic activities of MLEN in comparison with Diclofenac

While studying the antioxidant activity of plant extract, the plant extract demonstrates an initial increase in inhibition, peaking at 90.09% at 80 µg/ml, followed by a decline to 78.59% at 100 µg/ml, indicating a possible optimal concentration for effectiveness. While estimating the declining capacity of the plant extract, absorbance increases with plant extract concentration up to 80 µg/ml, indicating higher antioxidant or reactive activity. However, a drop in absorbance at 100 µg/ml suggests a non-linear response, possibly due to saturation or inhibitory effects.

Acknowledgement

The authors thank the Department of Botany and Microbiology, Acharya Nagarjuna University, Guntur, for providing necessary facilities for carrying out research.

References

1. Agrawal J. and Pal A., *Nyctanthes arbor-tristis* Linn—A critical ethnopharmacological review, *Journal of Ethnopharmacology*, **146**(3), 645-58 (2013)
2. Anashafer R.L., Harsha J., Selvin Pragalath Paul S.P. and Abbs Fen Reji T.F., Geometrical Electronic parameters, Vibrational Assignment docking studies, Antioxidant and Anticancer Activities of (1- benzyl)-2-(-2-Methoxyphenylamino thiazol-4yl)benzimidazole, *Res. J. Chem. Environ.*, **28**(4), 66-74 (2024)
3. Ashokkumar K., Dharshini M., Janani T., Shrravani Sri V. and Subhasidha R., *Nyctanthes arbor-tristis* Linn. (Night Jasmine): extraction techniques, phytochemical constituents and biological impacts of extracts and essential oil, *Future Journal of Pharmaceutical Sciences*, **10**(1), 117, <https://doi.org/10.1186/s43094-024-00694-2> (2024)

4. Audipudi A.V., Reddy P.H., Vajha M. and Bhaskar C.V., Evaluation of anti-arthritis and immunostimulatory activities of chloroform extracts of leaves of *Vitex negundo* L., *Int. J. Pharmacognosy and Phytochem. Res.*, **6**(2), 194-9 (2014)
5. Aurade G.K., Patil S.S., Shetkar M.A., Chandan Shive P.D. and Usnale S.V., Medicinal uses of plant *Nyctanthes arbor-tristis* Linn (Parijat): A Review, *Int. J. Pharmaceutical Res and Applications*, **7**(2), 47-56, doi: 10.35629/7781-07024756 (2022)
6. Azma W.A., Khan A.A. and Fazil M., Review on *Nyctanthes arbor-tristis* (harsinghar)-a herbal medicament with special reference to unani medicine, *Indian Journal of Unani Medicine*, **XIV**, 61-8, <https://doi.org/10.53390/ijum.v14i1.10> (2021)
7. Bhalakiya H. and Modi N.R., Traditional medicinal uses, phytochemical profile and pharmacological activities of *Nyctanthes arbor-tristis*, *RJLBPCS*, **5**(1), 1003-23, doi: 10.26479/2019.0502.76 (2019)
8. Bijauliya R.K., Kannoja P., Mishra P., Singh P.K. and Kannaujia R., Pharmacognostical and Physiochemical Study on the Leaves of *Nyctanthes arbor-tristis* Linn., *Journal of Drug Delivery and Therapeutics*, **11**(4), 30-4, <http://dx.doi.org/10.22270/jddt.v11i4.4914> (2021)
9. Biju T.S., Phytochemical, *in-vitro* antioxidant and *in-vitro* cytotoxicity analysis of *Nyctanthes arbor-tristis* ethyl acetate extract, *International Journal of Early Childhood Special Education*, **14**(5), doi:10.9756/intjecse/v14i5.8 (2022)
10. Chakraborty R. and De S.D., A Brief Overview on the Health Benefits of *Nyctanthes arbor-tristis* Linn-A Wonder of Mother Nature, *Indo Global Journal of Pharmaceutical Sciences*, **12**, 197-204, <http://doi.org/10.35652/IGJPS.2022.12024> (2022)
11. Devasree L.D., Binuramesh C. and Michael R.D., Immunostimulatory effect of water-soluble fraction of *Nyctanthes arbor-tristis* leaves on the immune response in *Oreochromis mossambicus* (Peters), *Aquaculture Research*, **45**(10), 1581-90, doi: 10.1111/are.12104 (2014)
12. Dewi N.K., Fakhrudin N. and Wahyuono S., A comprehensive review on the phytoconstituents and biological activities of *Nyctanthes arbor-tristis* L., *Journal of Applied Pharmaceutical Science*, **12**(8), 009-17, doi: 10.7324/JAPS.2022.120802 (2022)
13. Godse C.S., Tathed P.S., Talwalkar S.S., Vaidya R.A., Amonkar A.J., Vaidya A.B. and Vaidya A.D., Antiparasitic and disease-modifying activity of *Nyctanthes arbor-tristis* Linn. in malaria: An exploratory clinical study, *Journal of Ayurveda and Integrative Medicine*, **7**(4), 238-48, <http://dx.doi.org/10.1016/j.jaim.2016.08.003> (2016)
14. Gurib-Fakim A., Medicinal plants: traditions of yesterday and drugs of tomorrow, *Molecular aspects of Medicine*, **27**(1), 1-93, <https://doi.org/10.1016/j.mam.2005.07.008> (2006)
15. Haque M.M., Sultana N., Abedin S.M., Hossain N. and Kabir S.E., Fatty acid analysis, cytotoxicity, antimicrobial and antioxidant activities of different extracts of the flowers of *Nyctanthes arbor-tristis* L., *Bangladesh Journal of Scientific and Industrial Research*, **55**(3), 207-14, <https://doi.org/10.3329/bjsir.v55i3.49394> (2020)
16. Heendeniya S.N. et al, Therapeutic efficacy of *Nyctanthes arbor-tristis* flowers to inhibit proliferation of acute and chronic primary human leukemia cells, *Biomolecules*, **10**(2), 165, <https://doi.org/10.3390/biom10020165> (2020)
17. Jaiswal P. and Thakur M.K., The Investigation on Total Phenolic Content and *in Vitro* Antioxidant Potential of Different Plant Parts of *Nyctanthes arbor-tristis* (Night Jasmine), *Int. J. Pharm. Sci. Res.*, **8**, 3547-51, [http://dx.doi.org/10.13040/IJPSR.0975-8232.8\[8\].3547-51](http://dx.doi.org/10.13040/IJPSR.0975-8232.8[8].3547-51) (2017)
18. Kakoti B.B., Pradhan P., Borah S., Mahato K. and Kumar M., Analgesic and Anti-Inflammatory Activities of the Methanolic Stem Bark Extract of *Nyctanthes arbor-tristis* Linn., *BioMed Research International*, <https://doi.org/10.1155/2013/826295> (2013)
19. Kannan M., Singh A.R., Kumar T.A., Jegatheswari P. and Subburayalu S., Studies on immuno-bioactivities of *Nyctanthes arbor-tristis* (Oleaceae), *African Journal of Microbiology Research*, **1**(6), 88-91, doi.org/10.5897/ajmr.9000668 (2007)
20. Kaur J. and Kaushal S., Chemical analysis, antimicrobial and antioxidant activities of harsingar (*Nyctanthes arbor-tristis*) essential oil, *Journal of Essential Oil Bearing Plants*, **23**(2), 230-45, <https://doi.org/10.1080/0972060X.2020.1759458> (2020)
21. Khanapur M., Avadhanula R.K. and Setty O.H., *In vitro* antioxidant, antiproliferative and phytochemical study in different extracts of *Nyctanthes arbor-tristis* flowers, *BioMed Research International*, 291271, <https://doi.org/10.1155/2014/291271> (2014)
22. Kiran K. and Singh G., Comprehensive phytochemical analysis and quantification of flavonoids and phenolics in leaves of *Nyctanthes arbor-tristis* Linn., *International Journal of Pharmaceutical Science and Medicine*, **2**(3), 75-77, doi: 10.70199/ijpsm.2.3.75-77 (2024)
23. Lad H. and Bhatnagar D., Modulation of oxidative stress mediators in the liver of adjuvant induced arthritic rats by *Nyctanthes arbor tristis*, *Clinical Phyto Science*, **3**, 1-8, DOI 10.1186/s40816-016-0041-4 (2017)
24. Meghashri S. and Gopal S., Biochemical characterisation of radical scavenging polyphenols from *Nyctanthes arbor-tristis*, *Journal of Pharmacy and Bioallied Sciences*, **4**(4), 341-4, doi: 10.4103/0975-7406.103277 (2012)
25. Michael J.S., Kalirajan A., Padmalatha C. and Singh A.R., *In vitro* antioxidant evaluation and total phenolics of methanolic leaf extracts of *Nyctanthes arbor-tristis* L., *Chinese Journal of Natural Medicines*, **11**(5), 484-7, [https://doi.org/10.1016/S1875-5364\(13\)60088-6](https://doi.org/10.1016/S1875-5364(13)60088-6) (2013)
26. Mishra A.K., Tiwari K.N., Saini R., Chaurasia J.K. and Mishra S.K., Assessment of antioxidant potential in seed extracts of *Nyctanthes arbor-tristis* L. and phytochemical profiling by Gas Chromatography-Mass Spectrometry system, *Brazilian Journal of Pharmaceutical Sciences*, **58**, e21180, <https://doi.org/10.1590/s2175-97902022e21180> (2022)
27. Mousum S.A., Ahmed S., Gawali B., Kwatra M., Ahmed A. and Lahkar M., *Nyctanthes arbor-tristis* leaf extract ameliorates hyperlipidemia-and hyperglycemia-associated nephrotoxicity by

improving antioxidant and anti-inflammatory status in high-fat diet–streptozotocin-induced diabetic rats, *Inflammopharmacology*, **26**, 1415-28, <https://doi.org/10.1007/s10787-018-0497-6> (2018)

28. Nanu R., Raghuvver I., Chitme H.R. and Chandra R., Free radical scavenging activity of *Nyctanthesarbortristis* in streptozotocin-induced diabetic rats, *Indian Journal of Pharmaceutical Education and Research*, **44**, 288-94, <http://www.ijper.org/article/282> (2010)

29. Padhy M., A review on medicinal plants *Withaniasomnifera* and *Nyctanthesarbor-tristis*: boosting of immune system during SARS-CoV-2, *Lett. Appl. NanoBioScience*, **9**, 1538-46, <https://doi.org/10.33263/LIANBS94.15381546> (2020)

30. Parekh S. and Soni A., *Nyctanthesarbor-tristis*: Comprehensive review on its pharmacological, antioxidant and anticancer activities, *J Appl BiolBiotechnol*, **8**(1), 95-104, doi: 10.7324/JABB.2020.80116 (2020)

31. Patel S. and Gokhale M., Comparative Study of Antioxidant Activity of Ethanol and Aqueous Extracts of Different Parts of *Nyctanthesarbor-tristis* Linn, *Pharmacognosy Journal*, **8**(2), doi: 10.5530/pj.2016.2.3 (2016)

32. Paul A., Barman A.K. and Ray S., A Study on Antimicrobial Properties and Medicinal Value of *Adhatodavasica*, *Centella asiatica*, *Paederiafoetida*, *Nyctanthesarbor-tristis*, *Ocimum tenuiflorum*, *International Journal of Current Microbiology and Applied Science*, **7**(5), 1406-13 (2018)

33. Rajani K.S., Naik S.K., Satapathy S. and Mishra S., Phytochemical analysis of the leaf extracts of *Nyctanthesarbor-tristis*, *Plant Archives*, **21**(1), 1878-1881, DOI: <https://doi.org/10.51470/PLANTARCHIVES.2021.v21.no1.261> (2021)

34. Rangika B.S., Dayananda P.D. and Peiris D.C., Hypoglycemic and hypolipidemic activities of aqueous extract of flowers from *Nyctanthesarbor-tristis* L. in male mice, *BMC Complement Altern Med.*, **15**(1), 1–9, doi: 10.1186/s12906-015-0807-0 (2015)

35. Rathee J.S., Hassarajani S.A. and Chattopadhyay S., Antioxidant activity of *Nyctanthesarbor-tristis* leaf extract, *Food Chemistry*, **103**(4), 1350-7 (2007)

36. Rathore B., Paul B., Chaudhury B.P., Saxena A.K., Sahu A.P. and Gupta Y.K., Comparative studies of different organs of *Nyctanthesarbor-tristis* in modulation of cytokines in murine model of arthritis, *Biomedical and Environmental Sciences*, **20**(2), 154 (2007)

37. Saha R.K., Acharya S., Shovon S.S., Apu A.S. and Roy P., Biochemical investigation and biological evaluation of the methanolic extract of the leaves of *Nyctanthesarbor-tristis* *in vitro*, *Asian Pacific Journal of Tropical Biomedicine*, **2**(3), S1534-41, [https://doi.org/10.1016/S2221-1691\(12\)60449-3](https://doi.org/10.1016/S2221-1691(12)60449-3) (2012)

38. Sharma A., Goel A., Gupta N. and Sharma B., Phytochemicals from *Nyctanthesarbor-tristis* and their biomedical implications, *J.*

Med. Pharm. Allied Sci., **12**, 6012-20, doi: 10.55522/jmpasV12I4.5284 (2023)

39. Shrestha D.K., Sapkota B.K. and Sharma K.P., Assessment of Phytochemicals, Antioxidant Activity, Total Phenolic and Flavonoid Contents of Selected Nepalese Medicinal Plants, *Nepal Journal of Biotechnology*, **10**(2), 45-51, <https://doi.org/10.54796/njb.v10i2.239> (2022)

40. Singh A.K. and Solanki S., Antimicrobial activity of *Nyctanthesarbor-tristis* against *Staphylococcus aureus*, *Streptococcus pyogenes*, *Pseudomonas aeruginosa* and *Salmonella typhi*, *Int. Jr. Pharm. Sci. and Research*, **13**(7), 1000-9, [http://dx.doi.org/10.13040/IJPSR.0975-8232.13\(7\).1000-09](http://dx.doi.org/10.13040/IJPSR.0975-8232.13(7).1000-09) (2022)

41. Singh J., Singh A.P. and Singh A.P., *Nyctanthesarbor-tristis*: A comprehensive review, *World J. Curr. Med. Pharm. Res.*, **3**, 74-8, <https://doi.org/10.37022/wjcmpr.v3i4.181> (2021)

42. Siriwardena V.S. and Arambewela L.S., Determination of Volatile Constituents of the Essential Oil and Absolute of *Nyctanthesarbor-tristis* L. Flowers Grown in Sri Lanka, *Journal of Tropical Forestry and Environment*, **4**(2), <https://doi.org/10.31357/jtfe.v4i2.1853> (2014)

43. Srivastava R., Trivedi D., Shukla G. and Srivastava P., *Nyctanthesarbor-tristis*: A wonder Indian herbal drug needs healthcare attention, *Biomedical Journal of Scientific & Technical Research*, **5**(3), 1-4, DOI: 10.26717/BJSTR.2018.05.001199 (2018)

44. Tanimale D.R., Sahgal G., Sherly D.G., Yen F.L., Asem M., Selvam P.P. and Ng C.T., Antioxidant and anti-inflammatory properties of *Nyctanthesarbor-tristis* leaf extracts in high glucose-induced Sprague-Dawley rats, *NeuroQuantology*, **20**(8), 5236, doi:10.14704/nq.2022.20.8.nq44556 (2022)

45. Uroos M., Abbas Z., Sattar S., Umer N., Shabbir A. and Sharif A., *Nyctanthesarbor-tristis* Ameliorated FCA-Induced Experimental Arthritis: A Comparative Study among Different Extracts, *Evidence-Based Complementary and Alternative Medicine*, 4634853, <https://doi.org/10.1155/2017/4634853> (2017)

46. Vaidya V.V., Pradhan P.M. and Shinde M.A., Physicochemical standardisation and metal analysis of *Nyctanthesarbor-tristis* Linn, *World Journal of Pharmaceutical Research*, **6**(3), doi: 10.20959/wjpr20173-8017 (2017)

47. Vankar P.S., Antioxidant Activity of the Flower of *Nyctanthesarbor-tristis* L., *International Journal of Food Engineering*, **4**(8), <https://doi.org/10.2202/1556-3758.1437> (2008)

48. Verma N., Kaur J. and Bhatia A., Stimulation of acetylcholinesterase activity with *Nyctanthesarbor-tristis* leaves extract in the malathion-treated immunosuppressed mice, *International Journal of Environmental Studies*, **58**(5), 45-54 (2001).

(Received 22nd July 2025, accepted 10th September 2025)